

REVIEW ARTICLE

Recent insights into USP7: Construct, pathophysiology, and inhibitors

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Abstract

The ubiquitin-proteasome pathway (UPP) is essential for proteostasis and cellular homeostasis. Most of the human proteins are degraded through the UPP in which proteins should be tagged with a specific polyubiquitin chain in a sequential cascade of E1 ubiquitin (Ub)-activating enzymes, namely, E2 Ub-conjugating enzymes and E3 Ub ligases. Meanwhile, the ubiquitination process can be reversed by deubiquitinating enzymes (DUBs), which protect the target proteins from ubiquitination, and so far, around 100 DUBs have been reported to present in human cells. Ubiquitin-specific protease 7 (USP7) is a member of the DUBs family, which has been reported to play crucial role in the development of human tumors and diseases; however, the molecular mechanisms of disease and malignant tumor progression mediated by USP7 has not been fully elucidated. In addition, the therapeutic potential of USP7 in cancer treatment remains to be further explored. Therefore, this review begins with a review of the structure and function of USP7, and then focuses on the development of USP7 inhibitors and their potential applications in various human diseases.

Keywords: Degradation; Ubiquitin-proteasome pathway; Deubiquitinating enzymes; Ubiquitin specific protease 7; Molecule inhibitors

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1. Introduction

Ubiquitination is an essential and important post-translational modification process for most of the proteins, through which the proteins are covalently tagged with ubiquitin (Ub) molecules. Ub molecule, which is composed of 76 amino acids, is widely expressed, and the structure of the molecules is highly conserved across different species^[1]. At the consumption of ATP, Ub molecules are covalently conjugated to a certain lysine residue of the target protein, which is further catalyzed by the E1 Ub-activating enzymes (for activation), E2 Ub-conjugating enzymes (for conjugation), and E3 Ub ligases (for ligation)^[2]. It has been reported that there are around 2 Ub-activating enzymes (E1s), at least 38 Ub-conjugating enzymes (E2s), and more than 600 Ub ligases (E3s) in human, and the specific combinations of these enzymes determine the specificity of the protein ubiquitination process. Interestingly, Ub molecules contain seven lysine residues, namely K6, K11, K27, K29, K33, K48, and K63, which could be conjugated to other Ub molecules, providing diversity, and specificity for the ubiquitination

process of the substrate proteins^[3]. On the other hand, ubiquitination is a reversible and dynamic process, where Ub molecule chains can be cleaved away from the target proteins by certain deubiquitinating enzymes (DUBs), protecting the target protein from degradation^[4]. In brief, DUBs are a type of ubiquitin-specific proteases (USPs), which can induce the hydrolysis of Ub chains from the specific substrate proteins, leading to recycling of the Ub molecules, processing of the Ub precursors, and reversing of the Ub conjugation^[5,6]. In terms of biological functions, DUBs can regulate several cellular processes in humans, such as protein stabilization^[7], gene transcription^[7], signal transduction^[8,9], cell cycle progression, protein localization^[10], DNA damage response, kinase activation^[11,12], and many other functions.

Further, there are around 100 DUBs in human cells, which are able to reverse the ubiquitination reaction by removing the covalently attached Ub molecules from the substrates. Based on their structure and functional characteristics, DUBs can be divided into five subfamilies: (1) USPs, the largest subfamily; (2) Ub C-terminal hydrolase; (3) ovarian tumor; (4) Machado-Josephin disease protein; and (5) Jab1/Mov34/Mpr1-Pad1N-terminal+(JAMM)^[13]. USP7 is a cysteine protease, which belongs to the USP subfamily. It has been widely reported that USP7 interacts with the substrates to prevent ubiquitination and degradation of the target proteins^[14]. In this review, the recent research advances about USP7 in terms of its structure, pathological processes, and development as specific inhibitors are discussed.

2. The structure of the USP7

USP7, also known as herpes virus-associated ubiquitin-specific protease (HAUSP), has been demonstrated to participate in various pathological processes and diseases, including neurological disorders, metabolic disorders, immune dysfunction, and particularly carcinogenesis. In addition, USP7 is highly expressed in different types of cancers, including breast cancer^[15], medulloblastoma cancer^[16], T-cell leukemia^[17], ovarian cancer^[18], lung cancer^[19], and multiple myeloma (MM)^[20]. Furthermore, USP7 is closely related to malignant progression through its deubiquitinating activity in stabilizing the oncogene proteins, such as c-Maf and MafB transcription factors^[20], E3 Ub ligase human homolog of double minute 2 (HDM2) protein^[21], oncoprotein c-Myc, and enhancer of zeste homolog 2 (EZH2) protein^[22,23]. Interestingly, USP7 also has been shown to stabilize several tumor suppressor proteins, including p53 and phosphatase and tensin homolog (PTEN)^[10,24]. Due to the diversity of its substrates, it is difficult to determine whether USP7 functions as an

oncogene or as a tumor suppressor; therefore, it is essential to identify the specific substrate proteins, to understand better the roles of USP7 in specific clinical setting. Thus, the structural domains of USP7 and their effects on the USP7 activity, and the correlation between USP7 and intracellular signaling pathways, as well as the molecular mechanisms underlying the effects of USP7 on cancer and other diseases are further discussed in this article. Finally, the application of USP7-specific inhibitors and their potential applications in human diseases are included in this article as well.

2.1. The tumor necrosis factor-receptor associated factor (TRAF) domain

USP7 belongs to the UBP subfamily featured with an N-terminal TRAF-like domain^[25]. The TRAF domain consists of 160–165 amino acid residues, which can mediate substrate recognition and interaction^[26]. It is well-known that TRAF domain binds to both p53 and HDM2, thereby mediating their deubiquitination^[26,27]. However, the deletion of the N-terminal TRAF-like domain does not prevent the deubiquitination of p53 and HDM2 by USP7, suggesting the presence of a secondary binding site in USP7 with the substrate proteins. Later, this secondary binding motif in USP7 is believed to be the ubiquitin-like (UBL) domain^[27], indicating that there are different binding modes between USP7 and the substrates.

2.2. The UBL domain

In addition to the TRAF domain, UBL domain is another common structural domain to UBPs. Most of the proteins within the UBP subfamily have one or more UBL domains, including USP4, USP6, USP7, USP9X/Y, USP11, USP14, USP15, USP19, USP24, USP31, USP32, USP34, USP43, USP47, USP48, and USP52^[28]. Of note, the UBL domain exhibits different regulatory functions. For example, the UBL domain in USP4 has dual functions; it modulates USP4 localization to the proteasome to enhance its catalytic activity, and at the same time, the UBL domain can mimic Ub molecule and competes for Ub binding, subsequently causing a reduction in the ubiquitination of the substrate^[29]. Interestingly, USP7 has five UBL domains (UBL1/2/3/4/5) at the C-terminus spanning from 560 to 1050 amino acids, and they are organized in a UBL1/2-UBL3-UBL4/5 sequence. The last two of the UBL domains are found to modulate the catalytic activity of USP7, specifically, by promoting a conformational change of USP7 through interacting with the catalytic domain (CD), leading to binding and reshaping of the catalytic center^[25,29,30]. In contrast, in the absence of these UBL domains, USP7 loses its catalytic activity about 100-fold^[31], meanwhile the UBL4/5 domains can activate

USP7 through binding and rearranging the switching loop in the CD, leading to about 100-fold increase in USP7 enzymatic activity^[30]. In addition, this process can be increased with the help of allosteric activator guanosine 5'-monophosphate synthase (GMPS)^[32], where GMPS can bind to UBL1/2/3, subsequently increases the interaction between CDs and UBL4/5, thereby promoting the USP7 activity^[32]. This process suggests that the UBL domains are indispensable for USP7 to perform its deubiquitinating functions.

2.3. The CD

The CD, which is essential for the cleaving process of the polyubiquitin chains, in USP7 is composed of 208–560 amino acids. It should be noted that the proteolytic activity of USP7 is highly diverse and varies depending on the specific substrate proteins^[25]. The catalytic triad contains three types of amino acids, including cysteine (Cys), histidine (His), and asparagine (Asn)/aspartic acid (Asp) residues^[33], of which the Asn/Asp residue is important for polarization of His residue, which in return stabilizes the catalytic activity of DUBs^[28]. Moreover, the triad is rearranged in the presence of a covalently bound Ub aldehyde to generate a catalytically competent state of the CD^[34]. It is important to note that, in the absence of the UBL domains, the CD will lose almost all of its activity.

3. USP7 regulates diverse types of ubiquitination

DUBs can hydrolyze the polyubiquitin chain of target proteins, thereby protecting them from degradation or modulating their function or cellular localization, depending on the types of ubiquitination process^[35]. As discussed above, ubiquitination is a process that covalently attaches a Ub molecule to the specific lysine (K) residue on a substrate protein. In a Ub molecule, there is a highly conserved region, which is composed of the seven lysine residues, namely, K6, K11, K27, K29, K23, K48, and K63, which may lead to the diverse forms of Ub modification. Based on the Ub chain, Ub modifications could be classified into monoubiquitination (substrate protein tagged with a single Ub molecule) and polyubiquitination (substrate protein tagged with a multiple Ub molecule), and according to the Ub binding sites, ubiquitination are accordingly named K6-, K11-, K27-, K29-, K23-, K48-, and K63-linked ubiquitination^[36]. Importantly, USP7 can hydrolyze almost all types of Ub through monoubiquitination as well as K48- and K63-linked polyubiquitination^[10,37]. Furthermore, USP7 has been shown to markedly reduce the polyubiquitination levels of the substrate proteins, thereby exerting diverse functions^[20,38,39].

3.1. Targeting monoubiquitination by USP7

In monoubiquitination, a single Ub molecule attaches to substrate proteins. PTEN protein, one of the best-known tumor suppressors which is involved in a plethora of cancer development, is a typical protein that is subject to monoubiquitination. PTEN can be ubiquitinated in various ways, including monoubiquitination, K27-, K48-, and K63- linked polyubiquitination, to modulate its subcellular localization, aggregation, stability, and functionality^[40-42]. It is known that nuclear localization of PTEN is associated with monoubiquitination of PTEN at K289, which is indispensable for its nuclear import and tumor suppression^[43]. In addition, USP7 hydrolyzes monoubiquitination of PTEN, which leads to PTEN nuclear exclusion and loss of phosphatase activity, eventually promoting prostate cancer tumorigenesis. Another example is mothers against decapentaplegic homolog 3 (SMAD3) protein. The biological activity of SMAD3 depends on the monoubiquitination modification. USP7 interacts with SMAD3 and subsequently deubiquitinates its monoubiquitination level, thereby repressing cancer progression in p53-deficient lung cancer. Furthermore, USP7 also deubiquitinates monoubiquitination of other proteins including forkhead box protein O4, proliferating cell nuclear antigen, histone H2B, and others^[44-46].

3.2. Hydrolyzing K48-linked polyubiquitination by USP7

Compared with monoubiquitination, polyubiquitination is more common in protein modification, and there are seven types of polyubiquitination on thousands of proteins. At present, USP7 is believed to have the largest number of substrate proteins in terms of deubiquitination as shown in Table 1. It is known that intratumoral hypoxia can induce hypoxia-inducible factor 1- α (HIF-1 α) and subsequently promote tumorigenesis, metastasis, and treatment resistance. A previous study has revealed that USP7 can remove the K48-linked polyubiquitination by physically interacting with HIF-1 α , thereby increasing the protein stability and resulting in tumor cell epithelial-mesenchymal transition, metastasis, and tumor progression^[47].

In addition, ADP-ribosylation factor 4 (ARF4) is a small guanine nucleotide-binding protein, which belongs to Ras superfamily of small G proteins. Previous studies have demonstrated that ARF4 is an anti-apoptotic protein, which acts as a BAX inhibitor, and its function is indispensable in glioblastoma tumorigenesis^[52,115]. Interestingly, USP7 has been shown to be significantly upregulated in glioblastoma patient samples. Further study revealed that USP7 directly binds to ARF4, catalyzes the removal of the K48-linked

Table 1. List of identified USP7 substrates

Substrates	Ref	Substrates	Ref	Substrates	Ref	Substrates	Ref
AR	[48]	GATA1	[49]	NF-κB	[50]	SIRT1	[51]
ARF4	[52]	GMPS	[32]	NHE3	[53]	SOCS1	[54]
ARMC5	[55]	HDM2	[56]	NLRP3	[57]	Sox9	[58]
BCR-ABL	[38]	HIF-α	[47]	N-Myc	[59]	SPRTN	[60]
Bmi1	[61]	Histone H2B	[32]	NOTCH1	[62]	SUMO	[63]
Bub3	[64]	Histone H3	[65]	Nrf1	[66]	Tip60	[67]
Cdc25A	[68]	HLTF	[69]	p53	[24]	TPP1	[70]
CDK1	[71]	ICP0	[72]	PHF8	[15]	TRAF6	[73]
Chk1	[74]	IKKγ	[75]	PLK1	[76]	TRIM27	[77]
Clapsin	[76]	IRSs	[78]	Polymerase η	[45]	Ube2E1	[79]
c-MYC	[80]	JMJD3	[81]	POT1	[82]	UHRF1	[83]
CSB	[84]	KDM6B	[85]	PPARγ	[86]	UL35	[87]
DAF16	[88]	LANA	[89]	PRC1	[90]	UVSSA	[91]
DLC1	[92]	LSD1	[93]	PTEN	[10]	vIRF1	[94]
DNMT1	[95]	MAF	[20]	Rb	[96]	vIRF4	[97]
E1B	[98]	MCL1	[99]	RECQL4	[100]	WDR5	[101]
EBNA1	[102]	MDMX	[103]	RING1B	[104]	XPC	[105]
ERα	[106]	MLL2	[101]	RIP1	[98]	YAP	[107]
EZH2	[108]	Mule/ARF-BP1	[109]	RNF168	[110]	ZNF638	[111]
FOXO4	[44]	NEK2	[112]	RUNX2	[113]	β-catenin	[114]

polyubiquitinated chain from ARF4, and promotes malignant progression of glioblastoma. In addition, *in vivo* experiments revealed that treatment of USP7 specific inhibitor, P5091, can significantly suppress the growth of tumor models through ARF4 degradation^[52].

3.3. Hydrolyzing other polyubiquitination by USP7

The Maf proteins are critical factors in myelomagenesis, and it is an independent biomarker indicating poor prognosis in MM, a malignant cancer that is derived from plasma cells. In our previous work, we found that Maf proteins, including c-Maf and MafB, are substrates of USP7, indicated by affinity-purification couple tandem mass spectrometry analysis of c-Maf and MafB interactomes^[20]. In addition, USP7 can interact with Maf proteins through both TRAF and UBL domains, thereby preventing c-Maf and MafB from polyubiquitination and increasing their stability by prolonging their half-lives. Further, knockdown of USP7 or treatment with a specific small molecule inhibitor against USP7 leads to Maf protein degradation, followed by MM cell apoptosis^[20].

In addition, we also identified the fusion protein BCR-ABL, the fundamental non-receptor tyrosine kinase in chronic myelogenous leukemia (CML), which acts as a substrate of USP7^[38]. Cell-based experiments

demonstrated that USP7 specifically binds to BCR-ABL protein, subsequently protects them from proteasome degradation, resulting in CML cell proliferation. Further, overexpression of USP7 increases the protein stabilization of BCR-ABL and induces CML cell survival; in contrast, knockdown of USP7 decreases the protein level of BCR-ABL and its downstream target protein, phosphorylated STAT5, resulting in the inhibition of CML cell growth^[38].

4. The broad substrate spectrum of USP7

The synthesis and degradation of proteins are a dynamic equilibrium process, which maintains the homeostasis of the cells. In addition, protein degradation is an essential process, which is regulated through the UPP pathway, and this process can be reversed by the DUB. USP7 is a DUB with a long list of substrate proteins (oncoproteins or tumor suppressor proteins); therefore, USP7 is known to be widely involved in the development and progression of various cancers. Abnormal activation of an oncogene or dysfunction of a tumor suppressor gene in cells may result in malignancy and induction of a series of abnormal gene expression.

TP53, also known as p53, is a protein encoded by the *TP53* gene in human and plays an important role in tumor

suppression. The tumor suppressive role of p53 protein has been widely reported, and as a transcriptional factor, p53 regulates numerous physiology processes, such as cell cycle, DNA damage repair, cell apoptosis, antitumor activity, and cell stress response^[116]. In contrast, mutations and dysregulation of p53 have been reported to occur in more than 50% of all malignancies, leading to the loss of its antitumor activity^[117]. The stability of p53 is regulated through the UPP and its polyubiquitination, and mediated by HDM2, an E3 Ub ligase^[118]. Interestingly, HDM2 is also degraded through the UPP in an autoubiquitination pattern, at the same time p53 and HDM2 interacts with each other by forming a feedback loop^[119]. Interestingly, USP7 can regulate both p53 and mouse double minute 2 homolog (MDM2). USP7 interacts with p53 through its TRAF-like domain and stabilizes the p53 protein level by decreasing its ubiquitination level, even in the presence of HDM2^[24]. Meanwhile, USP7 also binds with HDM2 and inhibits its polyubiquitination^[21]. Thus, these three proteins form a complex, as evidenced by their crystal structure analyses, where both HDM2 and p53 interacts with the TRAF-like domain of USP7, further, the interaction of HDM2/USP7 is closer than p53/USP7^[120,121]. The previous studies have shown that partial knockdown of USP7 induces the degradation of p53, while the complete inhibition of USP7 expression leads to the stabilization of p53, and this phenomenon is related to HDM2^[21,122]. Another study reported that USP7 stabilizes both p53 and HDM2 protein *in vivo*. However, it is believed that USP7 maintains HDM2 to a certain protein level, to regulate the polyubiquitination degradation of p53.

In addition, USP7 has been shown to have other substrate proteins, including PTEN, c-Maf, MafB, transactivator of transcription, NAD-dependent deacetylase sirtuin-1, BCR-ABL, GATA-binding factor 1 (also known as GATA1), and ring finger protein 168^[10,20,21,24,38,49,51,110,123]. Many of these proteins are associated with cell cycle, DNA damage, regulation of transcription, and tumorigenesis. In addition, some of the substrate proteins have been shown to function as a mediator in certain signaling pathways, indicating the complexity of the substrate proteins function in human cells, thus, more studies are required to explore and understand its functions.

5. USP7 modulates signaling transduction

It has been reported that USP7 is involved in multiple cellular signaling pathways, such as PI3K/PTEN/AKT signaling^[10,124], Wnt/ β -catenin signaling^[39], Hippo pathway^[107], NF- κ B signaling^[125], type I IFN signaling^[126], and DNA damage signaling^[110] (Table 2). These pathways have different effects on cells, although their activities are regulated by USP7. For example, in multiple cellular

Table 2. Signaling pathway regulated by USP7

Signaling pathway	Ref	Signaling pathway	Ref
p300-p53/p21 pathway	[128]	BCR-ABL signaling	[38]
EZH2-CCF-cGAS signaling	[129]	MDM2/MDMX-p53 pathway	[130]
FBP1-DNMT1 pathway	[131]	NEDD4L-SMAD pathway	[132]
Cell cycle and EMT pathway	[133]	JMJD3/CLU signaling	[81]
p53/TfR1 pathway	[134]	Wnt/ β -catenin signaling	[39]
PI3K/Akt pathway	[124]	Hippo pathway	[107]
Akt/ERK signaling	[135]	NF- κ B signaling	[136]
AMPK pathway	[124]	Shh pathway	[16]
NOX4/NLRP3 pathway	[137]	Glucose Metabolism signaling	[138]
Sox9-PTHrP-PTH1R axis	[58]	Insulin/IGF-1-like signaling	[88]
HIF-1 α signaling pathway	[139]	Hedgehog signaling	[140]
Shoc2-ERK1/2 pathway	[141]	p53-MDM2 pathway	[21]

systems, USP7 directly interacts with and stabilizes Axin protein by decreasing its polyubiquitination and inhibiting the Wnt/ β -catenin pathway thereby suppressing Wnt-induced osteoblast differentiation^[39]. In addition, Hippo pathway is essential to the organ development, and its deregulation has been identified in a wide variety of tumors. Further, a transcription coactivator Yorkie has been demonstrated to be stabilized by USP7 in hepatocellular carcinoma (HCC) cells; interestingly, this protein is the upstream of the Hippo signaling^[107]. Taken together, USP7, which regulates Hippo pathway, could be a potential therapeutic target for HCC. In addition, the nuclear factor kappa light chain enhancer of activated B cells (NF- κ B) signal transduction pathway has been described previously to participate in the development and dysfunction of the immune system. Most of the proteins in this pathway are known to regulate biological processes, such as innate and adaptive immunity, inflammation, and stress responses. USP7 stabilizes NF- κ B signaling and increases its transcription, leading to the expression of downstream genes in response to the immune system activation^[127].

6. USP7 and diseases

6.1. The roles of USP7 in viral infection and inflammation

Interferon (IFN) is a low molecular glycoprotein induced by virus, bacteria, and IFN inducers acting on cells. IFN is not toxic to the cell itself, but it inhibits the replication of a variety of viruses and possesses antitumor activity^[142].

In general, IFN can be divided into three types: (1) type I IFN, including IFN- α (20 subtypes) and IFN- β (1 subtype), which are produced mainly by white blood cells, fibroblasts, and virus-infected tissue cells; (2) type II IFN, including IFN- γ , and it has only one subtype, which is produced by activated T cells and NK cells; and (3) type III IFN, including IFN- γ 1, IFN- γ 2, and IFN- γ 3. The main functions of type I IFN are inhibiting viral replication, providing resistance to parasitic infection, inhibiting cell proliferation, stimulating immune cells, eliciting antitumor activity, and regulating immunomodulatory system. Meanwhile, type II IFN possesses antiviral and anti-proliferation activities, and its main function is in immune regulation^[143,144]. For example, in response to virus invasion, the body produces IFN, and the process is mediated by several factors. USP7 has been reported previously as an important negative modulator of virus-induced signaling, where USP7 interacts with the E3 Ub ligase tripartite motif 27 (TRIM27), and their interaction is further enhanced after virus invasion. In addition, it has been reported that TNF receptor-associated factor family member-associated NF- κ B binding kinase (TBK)-1, which phosphorylates IFN regulatory factor (IRF)-3 and IRF-7, promotes the nuclear translocation and induces the expression of antiviral products^[145,146]. Interestingly, TBK-1 is degraded through the UPP, which is mediated by TRIM27. More remarkably, USP7 decreases TBK-1 expression and inhibits IFN antiviral efficacy by stabilizing TRIM27 protein^[126]. In another example, the suppressor of cytokine signaling 1 is a negative regulator of IFN-induced gene expression, which inhibits IFN antiviral efficacy by interacting with USP7 through its deubiquitinase activity^[54,147,148]. In addition, the p53 protein has been identified as a positive regulator in response to virus infection^[149]. Kaposi's sarcoma-associated herpesvirus (KSHV)-encoded viral interferon regulatory factor 4 (vIRF4), is one of the KSHV protein which interacts with USP7 and suppresses its enzyme activity, resulting in p53 degradation. Further studies have identified that vIRF4 stabilizes HDM2 by inhibiting its autoubiquitination process, and destabilizes p53^[97,150]. The above findings suggest the crucial role of USP7 in antiviral response; therefore, USP7 can be treated as a potential target for antiviral therapy. However, USP7 also plays an important role in homeostasis of normal cells; therefore, these factors should be considered when using USP7 inhibitors.

6.2. USP7 in diabetic foot

Diabetes mellitus (DM) is a common disease that happens worldwide. Blood vessel disorders are frequently seen in patients with DM, and it is commonly associated with

diabetic foot, one of the most common complications of a diabetic patients. However, at present, there is no effective treatment for diabetic foot. USP7 expression is found to be upregulated in human umbilical vein endothelial cells (HUVECs), when advanced glycation end products are used to establish the diabetic cell model^[151]. Inhibition of USP7 was shown to attenuate HUVECs cell cycle arrest and cell senescence. It has been further noted that the mechanism is regulated by USP7, which promotes cycle arrest and senescence of HUVECs cells by regulating the level of p53 polyubiquitination. The downregulation of p53 can reverse USP7-mediated HUVECs cell cycle arrest and cell senescence. Treatment of diabetic rat models with USP7 inhibitors can relieve the symptoms of diabetic foot. In conclusion, inhibition of USP7 induces p53 protein degradation and alleviates the symptoms of diabetic foot. These results showed that USP7 is a potential target for the treatment of diabetic foot ulcers.

6.3. USP7 in pulmonary hypertension

Pulmonary hypertension is a hemodynamic and pathophysiological condition in which the pulmonary artery pressure increases beyond a certain threshold. Pulmonary hypertension can be a group of independent diseases, complications, or syndromes. Pulmonary hypertension can be caused by the changes in the pulmonary artery itself, while others factors which can contribute to the development of this disease include genetic factors, drugs, toxins, and other diseases, such as congenital heart disease^[152,153].

Recent work has uncovered the role of USP7 in the development of pulmonary hypertension^[154]. Abnormal hyperplasia of pulmonary smooth muscle cells (PASCs) is an important pathological feature of pulmonary hypertension, and platelet-derived growth factor (PDGF)-induced PASCs proliferation is a critical factor in the development of pulmonary hypertension. Studies have revealed that PDGF increases the protein expression of USP7 and its downstream MDM2 and cyclinD1, resulting in the induction of PASCs proliferation. In addition, downregulation of USP7 can attenuate PDGF-induced MDM2, cyclinD1 stabilization, and cell proliferation. On the other hand, MG132 treatment can also abolish PDGF-induced cyclinD1 elevation and cell proliferation. In short, inhibition of USP7 activity might be a potential therapeutic strategy for the treatment of pulmonary hypertension. Notably, USP7 inhibits tumor progression in p53-deficient lung cancer cells, and USP7 maintains homeostasis in normal cells. Therefore, other factors should be considered when USP7 inhibitors are used for pulmonary hypertension.

6.4. USP7 in cancers

Among the diseases, cancers are the most intensively investigated research area in terms of USP7. USP7 is highly overexpressed in most tumors, such as breast cancer, medulloblastoma, T-cell leukemia, ovarian cancer, HCC, MM, prostate cancer, and CML^[15-17,20,23,38,107], suggesting the role of USP7 in tumor promotion. Relative protein expression in **Figure 1** shows USP7 expression in several cancers^[155].

USP7 is overexpressed in human breast cancer, and its expression is positively correlated with its substrate protein, PHF8. USP7 decreases the polyubiquitination of PHF8 and increases the expression of cyclin A2 protein, which is an essential and important protein for the development of breast cancer^[15]. Estrogen receptor α (ER α) is reported to be overexpressed in about 70% breast tumors. Studies have identified the positive correlation between ER α and USP7, depending on the deubiquitinase activity of USP7 and their interactions, where USP7 silencing led to cell growth inhibition and delayed tumor growth^[106]. In addition, USP7 has also been proven to be associated with poor prognosis of breast cancer, suggesting USP7 as a potential target for clinical treatment. Further, USP7 acts as a DUB of p53; therefore, the negative effects of p53 protein degradation should be considered when using USP7 inhibitors for breast cancer treatment.

Prostate cancer is a form of cancer that develops in the prostate, and is considered a slow-growing tumor. The incidence of prostate cancer is increasing by 2–3% every year, with more than 10% of death in male^[156]. As a histone methylase, EZH2 plays an important role in the development of diverse tumor, including prostate cancer. Studies identified that the deubiquitinase USP7 is highly expressed in prostate cancer, and more importantly, USP7 interacts with EZH2 and increases its protein stabilization through its deubiquitinating activity. In contrast, a reduction in USP7 expression fails to rescue the degradation of EZH2 through the UPP, resulting in the inhibition of cell growth and the suppression of tumor progression^[23]. Consistent with previous reports, USP7-specific inhibitors in combination with PARP inhibitor can produce a more lethal effect on prostate cancer cells, suggesting USP7 as a promising therapeutic target for prostate cancer.

HCC is one of the most common malignancies in China, and its incidence is ranked the third highest across the country^[157]. Studies have identified that USP7 is frequently overexpressed in HCC tissues, and positively correlated with poor prognosis, indicating UPP's role in the development of HCC^[158]. In addition, recent studies have demonstrated that USP7 promotes the proliferation, invasion, and migration of HCC cells through activating the bcl-2-like protein 4 (Bax), which subsequently

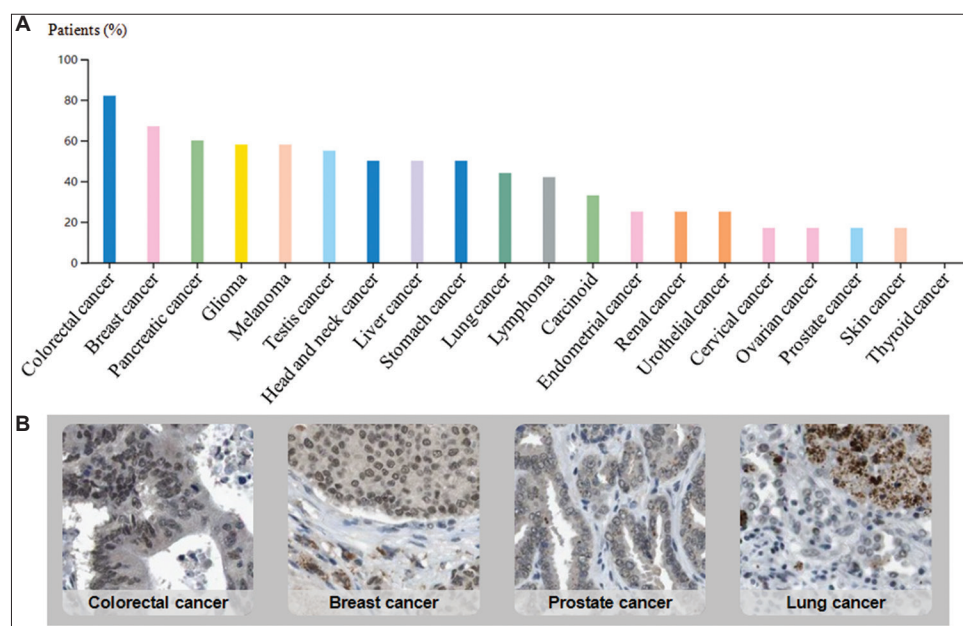


Figure 1. USP7 is highly expressed in cancer. (A) USP7 expression analysis of TCGA samples from The Human Protein Atlas database. Image reproduced from USP7 Protein Expression Summary; Malignant cells showed weak to moderate nuclear and cytoplasmic immunoreactivity (antibody CAB008108); available from <https://www.proteinatlas.org/ENSG00000187555-USP7/pathology> (image credit: The Human Protein Atlas). (B) Immunohistochemical staining of USP7 in representative tumors via the Human Protein Atlas (HPA) database. USP7 is highly expressed in a variety of tumors, including colorectal cancer, breast cancer, prostate cancer, and lung cancer. Image reproduced from USP7 Protein Expression; available from <https://www.proteinatlas.org/ENSG00000187555-USP7/pathology> (image credit: The Human Protein Atlas).

induces tumorigenesis and chemoresistance^[159]. On the other hand, USP7 stabilizes thyroid hormone receptor-interacting protein 12, which subsequently induces the polyubiquitination of p14(ARF), inactivate p14(ARF), and lastly promotes HCC progression^[160]. The above studies corroborate USP7 as the novel potential therapeutic target for HCC treatment.

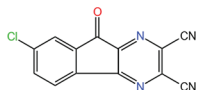
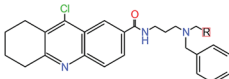
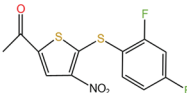
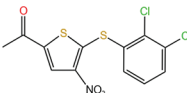
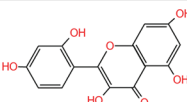
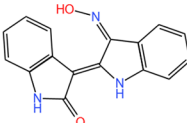
Hematological malignancies are usually caused by uncontrolled cell proliferation. MM is a cancer of blood plasma cells with high incidence and mortality rates, and currently there is no effective treatment strategy. MafB/c-Maf are highly expressed in several myeloma cell lines, such as RPMI-8226 and LP1, and they are critical transcription factors in myelomagenesis, promoting the cell growth, proliferation, metastasis, and cell cycle progression^[161]. USP7 has been shown to participate in MM progression by increasing the stabilization of MafB/c-Maf proteins. In addition, CML is a clonal malignancy of hematopoietic stem cells, featured with fusion protein kinase BCR-ABL. It has been demonstrated that deubiquitinase USP7 binds to BCR-ABL and reduces its polyubiquitination, thereby protecting it from degradation and inducing CML cell proliferation^[38]. Therefore, inhibition of USP7 kinase

activity could serve as a potential clinical treatment strategy for hematological disease.

7. Development of USP7-specific inhibitors

Given the diverse roles of USP7 in various diseases, USP7 has been widely believed to be an ideal therapeutic target, and many inhibitors against USP7 have been developed. The structural characteristic of USP7 has been described earlier in this article. The TRAF-like domain of USP7 is important for substrate binding. According to the role of TRAF domain in USP7, researchers have found the first inhibitor of USP7 through high-throughput screening (Table 3). Studies have revealed that HBX41108 disrupts the interaction between USP7 and HDM2, which inhibits USP7 deubiquitinating activity and fails to stabilize HDM2 protein, thereby inducing p53 accumulation and resulting in cell cycle arrest and apoptosis in tumor cells^[162]. Since USP7 belongs to the Cys protease family, which are highly conserved in terms of the structural composition, such as USP5, USP8, USP10 and CYLD, it has been demonstrated that both of their activity can be inhibited by HBX41108^[163,164], suggesting that the inhibition effect of HBX41108 on USP7 activity is not specific enough and further research is needed.

Table 3. The chemical structure of USP7 inhibitors, their validated targets, their effect on cell level and tumor model growth

Name	Structural formula	Validated targets	Functional consequence	<i>In vitro</i> cell suppression	Ref
HBX 41108		↓MDM2↑p53; p21	Cell cycle arrest Apoptosis	Colon cancer cells	[162]
HBX 19818		↓MDM2↑p53; p21	Cell cycle arrest Apoptosis Growth inhibition	Colon cancer cells	[164]
P22077		↓MDM2; caspin; Pchk1; DDB1; TIP60↑p53; p21; Cle-Caspase-3	Growth inhibition Apoptosis	Colon cancer cells; Neuroblastoma cells; Hepatocellular cells; Lung cancer cells	[67,159,165-167]
P5091		↓MDM2; MDMX; MafB; c-Maf; BCR-ABL; EZH2↑p53; p21	Cell cycle arrest Apoptosis Autophagy growth inhibition	Colon cancer cells; Neuroblastoma cells; Hepatocellular cells; Glioblastoma cells; Prostate cancer cells; Ovarian cancer cells Multiple myeloma cells	[18,20,52,161,168]
Morin		↓ Prickle1, mTORC2	Cell migration inhibition	Rheumatoid arthritis cells	[169]
I3MO		↓ NEK2, PA28γ, PA200 NF-κB signaling	Apoptosis DNA damage Cell cycle arrest	Multiple myeloma cells	[170]

In addition, CD plays a key role in cleaving polyubiquitin chains, especially the Cys223 residue, which is essential for the deubiquitinase activity of USP7. Using biochemical assays and activity-based protein profiling in living systems, HBX19818 and HBX28258 were found to be catalytic activity inhibitors of USP7, which can selectively inhibit the USP7 deubiquitinase activity by binding to its catalytic active site^[165] (Table 3). In addition, these two compounds have a higher selectivity to USP7, and have been shown to decrease its downstream proteins, leading to cell cycle arrest and growth inhibition in several tumor cells^[164].

Two new compounds, P22077 and P5091, which were discovered through a high-throughput screen, have been shown to selectively inhibit USP7 activity^[161,165]. P22077 can activate p53 and its downstream protein p21 in human colon carcinoma cells. Further studies found that P22077 stabilizes p53 by inducing the degradation of HDM2^[57].

P5091, the most recognized inhibitor against USP7 activity, has been identified to induce MM cell apoptosis and inhibit xenograft tumor growth. Mechanistically, P5091 leads to the decrease of HDM2, and subsequently stabilizes p53 and p21 in tumor cells by inhibiting USP7 activity. Interestingly, when MM cell line KMS11 was treated with P5091, there was no obvious increase of p53, suggesting that p53 is dispensable for the cytotoxicity of P5091^[57]. It's worth pointing out that P5091 shows high specificity without affecting other protease activity, except for USP7^[161]. These findings suggest that the USP7 inhibitors have potential value in clinical therapy.

Recent studies have found that Morin (3, 5, 7, 2', 4'-pentahydroxyflavone), an anti-arthritis compound that is present in foods of plant origin, suppresses the pathological migration of fibroblast-like synoviocytes, and prevents focal adhesion turnover^[169]. A molecular operating environment software has been used to show that Morin could bind closely to His461, Met292, and Phe409 amino acid residues in the CD of USP7 through hydrogen bonds, thus hindering the rotation of the Phe409 side chain and subsequently inhibiting the binding of USP7 and substrate Ub. Further, USP7 protects Prickle1 from degradation, and this process can be reversed by Morin to block FLS migration.

Furthermore, indirubin-3'-monoxime (I3MO), is one of the derivatives of indirubin. The previous study has reported that the anti-MM activity of I3MO in both drug-sensitive and drug-resistant MM cells by sensitizing MM cells to bortezomib-induced apoptosis^[170]. In return, I3MO suppresses the growth of MM cells via down-regulating the USP7 expression, inducing NEK2 degradation, and suppressing NF- κ B signaling in MM.

In summary, exploring the potential inhibitors against USP7 can provide a novel therapeutic strategy in tumor treatment.

8. Conclusions

Ubiquitination and deubiquitination are two opposite biological processes, which work together to regulate the post-translational modification of proteins. Various biological functions of USP7 have been discovered, including its role as a vital regulator of transcription^[20], nuclear export^[10], inflammatory responses^[57], DNA damage repair^[11], antiviral responses^[54], and induction of cell apoptosis^[20,171]. Furthermore, USP7 is considered a cancer-promoting protein in a variety of tumors, including lung cancer, prostate cancer, HCC, as well MM. The progression of all the above diseases can be related to the substrate proteins of USP7. USP7 binds to its substrates and stabilizes their proteins or maintains their activity, thereby inducing abnormal cell proliferation and tumor development. Moreover, studies have uncovered that USP7 interacts with HDM2, which is a critical negative regulator of p53, to stabilize its protein level^[24]. Contrary to the oncogenic role of USP7, several studies have found that it has dual function in cancer development. For example, in colon carcinoma, USP7 inhibits cell proliferation *in vitro* and tumor growth *in vivo* in the presence of stress due to the constitutive expression level of p53 protein^[172]. Therefore, further studies are required to better understand the role of USP7 in tumors. More remarkably, in recent years, accumulating research has focused on designing small molecule inhibitors of USP7, mainly based on its physical structure or enzyme activity, to provide potential strategies for cancer treatment. However, the premise is we need to fully understand the role or mechanism of USP7 in tumor development, since USP7 stabilizes p53 and its function is unclear in the presence of stress or radiation damage^[172]. The diversity of USP7 substrates determines the complexity of its function in different diseases and tumors, and the complications of diseases bring new challenges for USP7 as a target in cancer therapy. Therefore, further investigation is required to evaluate the benefits and adverse effects of inhibiting the enzymatic activity of USP7. In addition, with the emergence of new molecular tools and in-depth exploration of USP7 substrates and different strategies to modulate USP7 activity, the therapeutic potential of USP7 in diseases will be further evaluated.

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Conflict of interest

The authors declare that they have no competing interests.

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References

1. Nakamura N, 2018, Ubiquitin system. *Int J Mol Sci*, 19(4): 1080.
<https://doi.org/10.3390/ijms19041080>
2. Hershko A, Heller H, Elias S, *et al.*, 1983, Components of ubiquitin-protein ligase system. Resolution, affinity purification, and role in protein breakdown. *J Biol Chem*, 258(13): 8206–8214.
3. Xu P, Duong DM, Seyfried NT, *et al.*, 2009, Quantitative proteomics reveals the function of unconventional ubiquitin chains in proteasomal degradation. *Cell*, 137(1): 133–145.
<https://doi.org/10.1016/j.cell.2009.01.041>
4. Nijman SM, Luna-Vargas MP, Velds A, *et al.*, 2005, A genomic and functional inventory of deubiquitinating enzymes. *Cell*, 123(5): 773–786.
<https://doi.org/10.1016/j.cell.2005.11.007>
5. Poondla N, Chandrasekaran AP, Kim KS, *et al.*, 2019, Deubiquitinating enzymes as cancer biomarkers: New therapeutic opportunities? *BMB Rep*, 52(3): 181–189.
<https://doi.org/10.5483/BMBRep.2019.52.3.048>
6. Pozhidaeva A, Bezsonova I, 2019, USP7: Structure, substrate specificity, and inhibition. *DNA Repair (Amst)*, 76: 30–39.
<https://doi.org/10.1016/j.dnarep.2019.02.005>
7. Wang S, Juan J, Zhang Z, *et al.*, 2017, Inhibition of the deubiquitinase USP5 leads to c-Maf protein degradation and myeloma cell apoptosis. *Cell Death Dis*, 8(9): e3058.
<https://doi.org/10.1038/cddis.2017.450>
8. Zhang C, Chen Y, Gan X, *et al.*, 2017, SAK-HV decreases the self-ubiquitination of MEKK1 to promote macrophage proliferation via MAPK/ERK and JNK Pathways. *Int J Mol Sci*, 18(4): 835.
<https://doi.org/10.3390/ijms18040835>
9. Lork M, Verhelst K, Beyaert R, 2017, CYLD, A20 and OTULIN deubiquitinases in NF- κ B signaling and cell death: So similar, yet so different. *Cell Death Differ*, 24(7): 1172–1183.
<https://doi.org/10.1038/cdd.2017.46>
10. Song MS, Salmena L, Carracedo A, *et al.*, 2008, The deubiquitylation and localization of PTEN are regulated by a HAUSP-PML network. *Nature*, 455(7214): 813–817.
<https://doi.org/10.1038/nature07290>
11. Jacq X, Kemp M, Martin NM, *et al.*, 2013, Deubiquitylating enzymes and DNA damage response pathways. *Cell Biochem Biophys*, 67(1): 25–43.
<https://doi.org/10.1007/s12013-013-9635-3>
12. Mathien S, Dél  ris P, Soulez M, *et al.*, 2017, Deubiquitinating enzyme USP20 regulates extracellular signal-regulated kinase 3 stability and biological activity. *Mol Cell Biol*, 37(9): e00432–16.
<https://doi.org/10.1128/MCB.00432-16>
13. Kim SY, Baek KH, 2019, TGF- β signaling pathway mediated by deubiquitinating enzymes. *Cell Mol Life Sci*, 76(4): 653–665.
<https://doi.org/10.1007/s00018-018-2949-y>
14. Sun T, Liu Z, Yang Q, 2020, The role of ubiquitination and deubiquitination in cancer metabolism. *Mol Cancer*, 19(1): 146.
15. Wang Q, Ma S, Song N, *et al.*, 2016, Stabilization of histone demethylase PHF8 by USP7 promotes breast carcinogenesis. *J Clin Invest*, 126(6): 2205–2220.
<https://doi.org/10.1172/JCI85747>
16. Zhan M, Sun X, Liu J, *et al.*, 2017, Usp7 promotes medulloblastoma cell survival and metastasis by activating Shh pathway. *Biochem Biophys Res Commun*, 484(2): 429–434.
<https://doi.org/10.1016/j.bbrc.2017.01.144>
17. Jin Q, Martinez CA, Arcipowski KM, *et al.*, 2019, USP7 cooperates with NOTCH1 to drive the oncogenic transcriptional program in T-cell leukemia. *Clin Cancer Res*, 25(1): 222–239.
<https://doi.org/10.1158/1078-0432.CCR-18-1740>
18. Wang M, Zhang Y, Wang T, *et al.*, 2017, The USP7 inhibitor P5091 induces cell death in ovarian cancers with different P53 status. *Cell Physiol Biochem*, 43(5): 1755–1766.
<https://doi.org/10.1159/000484062>
19. Zhao GY, Lin ZW, Lu CL, *et al.*, 2015, USP7 overexpression predicts a poor prognosis in lung squamous cell carcinoma and large cell carcinoma. *Tumour Biol*, 36(3): 1721–1729.
<https://doi.org/10.1007/s13277-014-2773-4>
20. He Y, Wang S, Tong J, *et al.*, 2020, The deubiquitinase USP7 stabilizes maf proteins to promote myeloma cell survival. *J Biol Chem*, 295(7): 2084–2096.
<https://doi.org/10.1074/jbc.RA119.010724>
21. Li M, Brooks CL, Kon N, *et al.*, 2004, A dynamic role of HAUSP in the p53-Mdm2 pathway. *Mol Cell*, 13(6): 879–886.
[https://doi.org/10.1016/s1097-2765\(04\)00157-1](https://doi.org/10.1016/s1097-2765(04)00157-1)
22. Nicklas S, Hillje AL, Okawa S, *et al.*, 2019, A complex of the ubiquitin ligase TRIM32 and the deubiquitinase USP7

- balances the level of c-Myc ubiquitination and thereby determines neural stem cell fate specification. *Cell Death Differ*, 26(4): 728–740.
<https://doi.org/10.1038/s41418-018-0144-1>
23. Zheng N, Chu M, Lin M, *et al.*, 2020, USP7 stabilizes EZH2 and enhances cancer malignant progression. *Am J Cancer Res*, 10(1): 299–313.
24. Li M, Chen D, Shiloh A, *et al.*, 2002, Deubiquitination of p53 by HAUSP is an important pathway for p53 stabilization. *Nature*, 416(6881): 648–653.
<https://doi.org/10.1038/nature737>
25. Tavana O, Gu W, 2017, Modulation of the p53/MDM2 interplay by HAUSP inhibitors. *J Mol Cell Biol*, 9(1): 45–52.
<https://doi.org/10.1093/jmcb/mjw049>
26. Brooks CL, Li M, Hu M, *et al.*, 2007, The p53--Mdm2--HAUSP complex is involved in p53 stabilization by HAUSP. *Oncogene*, 26(51): 7262–7266.
<https://doi.org/10.1038/sj.onc.1210531>
27. Ma J, Martin JD, Xue Y, *et al.*, 2010, C-terminal region of USP7/HAUSP is critical for deubiquitination activity and contains a second mdm2/p53 binding site. *Arch Biochem Biophys*, 503(2): 207–212.
<https://doi.org/10.1016/j.abb.2010.08.020>
28. Komander D, Clague MJ, Urbé S, 2009, Breaking the chains: Structure and function of the deubiquitinases. *Nat Rev Mol Cell Biol*, 10(8): 550–563.
<https://doi.org/10.1038/nrm2731>
29. Faesen AC, Luna-Vargas MP, Sixma TK, 2012, The role of UBL domains in ubiquitin-specific proteases. *Biochem Soc Trans*, 40(3): 539–545.
<https://doi.org/10.1042/BST20120004>
30. Faesen AC, Dirac AM, Shanmugham A, *et al.*, 2011, Mechanism of USP7/HAUSP activation by its C-terminal ubiquitin-like domain and allosteric regulation by GMP-synthetase. *Mol Cell*, 44(1): 147–159.
<https://doi.org/10.1016/j.molcel.2011.06.034>
31. Kim RQ, van Dijk WJ, Sixma TK, 2016, Structure of USP7 catalytic domain and three Ubl-domains reveals a connector α -helix with regulatory role. *J Struct Biol*, 195(1): 11–18.
<https://doi.org/10.1016/j.jsb.2016.05.005>
32. van der Knaap JA, Kumar BR, Moshkin YM, *et al.*, 2005, GMP synthetase stimulates histone H2B deubiquitylation by the epigenetic silencer USP7. *Mol Cell*, 17(5): 695–707.
<https://doi.org/10.1016/j.molcel.2005.02.013>
33. Cotto-Rios XM, Békés M, Chapman J, *et al.*, 2012, Deubiquitinases as a signaling target of oxidative stress. *Cell Rep*, 2(6): 1475–1484.
<https://doi.org/10.1016/j.celrep.2012.11.011>
34. Hu M, Li P, Li M, *et al.*, 2002, Crystal structure of a UBP-family deubiquitinating enzyme in isolation and in complex with ubiquitin aldehyde. *Cell*, 111(7): 1041–1054.
[https://doi.org/10.1016/s0092-8674\(02\)01199-6](https://doi.org/10.1016/s0092-8674(02)01199-6)
35. Culig Z, Santer FR, 2014, Androgen receptor signaling in prostate cancer. *Cancer Metastasis Rev*, 33(2-3): 413–427.
<https://doi.org/10.1007/s10555-013-9474-0>
36. Swatek KN, Komander D, 2016, Ubiquitin modifications. *Cell Res*, 26(4): 399–422.
<https://doi.org/10.1038/cr.2016.39>
37. Huang YT, Cheng AC, Tang HC, *et al.*, 2021, USP7 facilitates SMAD3 autoregulation to repress cancer progression in p53-deficient lung cancer. *Cell Death Dis*, 12(10): 880.
<https://doi.org/10.1038/s41419-021-04176-8>
38. Jiang S, Wang X, He Y, *et al.*, 2021, Suppression of USP7 induces BCR-ABL degradation and chronic myelogenous leukemia cell apoptosis. *Cell Death Dis*, 12(5):456.
<https://doi.org/10.1038/s41419-021-03732-6>
39. Ji L, Lu B, Zamponi R, *et al.*, 2019, USP7 inhibits Wnt/ β -catenin signaling through promoting stabilization of Axin. *Nat Commun*, 10(1): 4184.
<https://doi.org/10.1038/s41467-019-12143-3>
40. Lee YR, Chen M, Lee JD, *et al.*, 2019, Reactivation of PTEN tumor suppressor for cancer treatment through inhibition of a MYC-WWPI inhibitory pathway. *Science*, 364(6441): eaau0159.
<https://doi.org/10.1126/science.aau0159>
41. Wang G, Zhuang Z, Shen S, *et al.*, 2022, Regulation of PTEN and ovarian cancer progression by an E3 ubiquitin ligase RBCK1. *Hum Cell*, 35(3): 896–908.
<https://doi.org/10.1007/s13577-022-00681-w>
42. He YM, Zhou XM, Jiang SY, *et al.*, 2022, TRIM25 activates AKT/mTOR by inhibiting PTEN via K63-linked polyubiquitination in non-small cell lung cancer. *Acta Pharmacol Sin*, 43(3): 681–691.
<https://doi.org/10.1038/s41401-021-00662-z>
43. Trotman LC, Wang X, Alimonti A, *et al.*, 2007, Ubiquitination regulates PTEN nuclear import and tumor suppression. *Cell*, 128(1): 141–156.
<https://doi.org/10.1016/j.cell.2006.11.040>
44. van der Horst A, de Vries-Smits AM, Brenkman AB, *et al.*, 2006, FOXO4 transcriptional activity is regulated by monoubiquitination and USP7/HAUSP. *Nat Cell Biol*, 8(10): 1064–1073.
<https://doi.org/10.1038/ncb1469>
45. Qian J, Pentz K, Zhu Q, *et al.*, 2015, USP7 modulates UV-induced PCNA monoubiquitination by regulating DNA

- polymerase eta stability. *Oncogene*, 34(36): 4791–4796.
<https://doi.org/10.1038/onc.2014.394>
46. Sarkari F, Sanchez-Alcaraz T, Wang S, *et al.*, 2009, EBNA1-mediated recruitment of a histone H2B deubiquitylating complex to the Epstein-Barr virus latent origin of DNA replication. *PLoS Pathog*, 5(10): e1000624.
47. Wu HT, Kuo YC, Hung JJ, *et al.*, 2016, K63-polyubiquitinated HAUSP deubiquitinates HIF-1 α and dictates H3K56 acetylation promoting hypoxia-induced tumour progression. *Nat Commun*, 7: 13644.
<https://doi.org/10.1038/ncomms13644>
48. Chen ST, Okada M, Nakato R, *et al.*, 2015, The deubiquitinating enzyme USP7 regulates androgen receptor activity by modulating its binding to chromatin. *J Biol Chem*, 290(35): 21713–21723.
<https://doi.org/10.1074/jbc.M114.628255>
49. Liang L, Peng Y, Zhang J, *et al.*, 2019, Deubiquitylase USP7 regulates human terminal erythroid differentiation by stabilizing GATA1. *Haematologica*, 104(11): 2178–2187.
50. Mitxitorena I, Somma D, Mitchell JP, *et al.*, 2020, The deubiquitinase USP7 uses a distinct ubiquitin-like domain to deubiquitinate NF- κ B subunits. *J Biol Chem*, 295(33): 11754–11763.
<https://doi.org/10.1074/jbc.RA120.014113>
51. Song N, Cao C, Tian S, *et al.*, 2020, USP7 Deubiquitinates and stabilizes SIRT1. *Anat Rec (Hoboken)*, 303(5): 1337–1345.
<https://doi.org/10.1002/ar.24252>
52. Pan T, Li X, Li Y, *et al.*, 2021, USP7 inhibition induces apoptosis in glioblastoma by enhancing ubiquitination of ARF4. *Cancer Cell Int*, 21(1): 508.
<https://doi.org/10.1186/s12935-021-02208-z>
53. Han Y, Yun CC, 2020, Ubiquitin-specific peptidase 7 (USP7) and USP10 mediate deubiquitination of human NHE3 regulating its expression and activity. *Faseb J*, 34(12): 16476–16488.
<https://doi.org/10.1096/fj.202001875R>
54. Yuan Y, Miao Y, Zeng C, *et al.*, 2020, Small-molecule inhibitors of ubiquitin-specific protease 7 enhance Type-I interferon antiviral efficacy by destabilizing SOCS1. *Immunology*, 159(3): 309–321.
<https://doi.org/10.1111/imm.13147>
55. Yan G, Liu N, Tian J, *et al.*, 2021, Deubiquitylation and stabilization of ARMC5 by ubiquitin-specific processing protease 7 (USP7) are critical for RCC proliferation. *J Cell Mol Med*, 25(6): 3149–3159.
<https://doi.org/10.1111/jcmm.16306>
56. Meulmeester E, Maurice MM, Boutell C, *et al.*, 2005, Loss of HAUSP-mediated deubiquitination contributes to DNA damage-induced destabilization of Hdmx and Hdm2. *Mol Cell*, 18(5): 565–576.
<https://doi.org/10.1016/j.molcel.2005.04.024>
57. Palazón-Riquelme P, Worboys JD, Green J, *et al.*, 2018, USP7 and USP47 deubiquitinases regulate NLRP3 inflammasome activation. *EMBO Rep*, 19(10): e44766.
<https://doi.org/10.15252/embr.201744766>
58. Dong X, Xu X, Yang C, *et al.*, 2021, USP7 regulates the proliferation and differentiation of ATDC5 cells through the Sox9-PTHrP-PTH1R axis. *Bone*, 143: 115714.
<https://doi.org/10.1016/j.bone.2020.115714>
59. Tavana O, Li D, Dai C, *et al.*, 2016, HAUSP deubiquitinates and stabilizes N-Myc in neuroblastoma. *Nat Med*, 22(10): 1180–1186.
<https://doi.org/10.1038/nm.4180>
60. Perry M, Biegert M, Kollala SS, *et al.*, 2021, USP11 mediates repair of DNA-protein cross-links by deubiquitinating SPRTN metalloprotease. *J Biol Chem*, 296: 100396.
<https://doi.org/10.1016/j.jbc.2021.100396>
61. Nguyen LK, Muñoz-García J, Maccario H, *et al.*, 2011, Switches, excitable responses and oscillations in the Ring1B/Bmi1 ubiquitination system. *PLoS Comput Biol*, 7(12): e1002317.
<https://doi.org/10.1371/journal.pcbi.1002317>
62. Shan H, Li X, Xiao X, *et al.*, 2018, USP7 deubiquitinates and stabilizes NOTCH1 in T-cell acute lymphoblastic leukemia. *Signal Transduct Target Ther*, 3: 29.
<https://doi.org/10.1038/s41392-018-0028-3>
63. Lecona E, Rodriguez-Acebes S, Specks J, *et al.*, 2016, USP7 is a SUMO deubiquitinase essential for DNA replication. *Nat Struct Mol Biol*, 23(4): 270–277.
<https://doi.org/10.1038/nsmb.3185>
64. Giovinazzi S, Sirleto P, Aksenova V, *et al.*, 2014, Usp7 protects genomic stability by regulating Bub3. *Oncotarget*, 5(11): 3728–3742.
<https://doi.org/10.18632/oncotarget.1989>
65. Yamaguchi L, Nishiyama A, Misaki T, *et al.*, 2017, Usp7-dependent histone H3 deubiquitylation regulates maintenance of DNA methylation. *Sci Rep*, 7(1): 55.
<https://doi.org/10.1038/s41598-017-00136-5>
66. Han JJW, Ho DV, Kim HM, *et al.*, 2021, The deubiquitinating enzyme USP7 regulates the transcription factor Nrf1 by modulating its stability in response to toxic metal exposure. *J Biol Chem*, 296: 100732.
<https://doi.org/10.1016/j.jbc.2021.100732>
67. Dar A, Shibata E, Dutta A, 2013, Deubiquitination of Tip60 by USP7 determines the activity of the p53-dependent

- apoptotic pathway. *Mol Cell Biol*, 33(16): 3309–3320.
<https://doi.org/10.1128/MCB.00358-13>
68. Das S, Chandrasekaran AP, Jo KS, *et al.*, 2020, HAUSP stabilizes Cdc25A and protects cervical cancer cells from DNA damage response. *Biochim Biophys Acta Mol Cell Res*, 1867(12): 118835.
<https://doi.org/10.1016/j.bbamcr.2020.118835>
69. Qing P, Han L, Bin L, *et al.*, 2011, USP7 regulates the stability and function of HLTF through deubiquitination. *J Cell Biochem*, 112(12): 3856–3862.
<https://doi.org/10.1002/jcb.23317>
70. Zemp I, Lingner J, 2014, The shelterin component TPP1 is a binding partner and substrate for the deubiquitinating enzyme USP7. *J Biol Chem*, 289(41): 28595–28606.
<https://doi.org/10.1074/jbc.M114.596056>
71. Galarreta A, Valledor P, Ubieto-Capella P, *et al.*, 2021, USP7 limits CDK1 activity throughout the cell cycle. *Embo J*, 40(11): e99692.
<https://doi.org/10.15252/embj.201899692>
72. Holowaty MN, Sheng Y, Nguyen T, *et al.*, 2003, Protein interaction domains of the ubiquitin-specific protease, USP7/HAUSP. *J Biol Chem*, 278(48): 47753–47761.
<https://doi.org/10.1074/jbc.M307200200>
73. Zhao XB, Ji FY, Li HR, *et al.*, 2020, P22077 inhibits LPS-induced inflammatory response by promoting K48-linked ubiquitination and degradation of TRAF6. *Aging (Albany NY)*, 12(11): 10969–10982.
<https://doi.org/10.18632/aging.103309>
74. Alonso-de Vega I, Martín Y, Smits VA, 2014, USP7 controls Chk1 protein stability by direct deubiquitination. *Cell Cycle*, 13(24): 3921–3926.
<https://doi.org/10.4161/15384101.2014.973324>
75. Daubeuf S, Singh D, Tan Y, *et al.*, 2009, HSV ICP0 recruits USP7 to modulate TLR-mediated innate response. *Blood*, 113(14): 3264–3275.
<https://doi.org/10.1182/blood-2008-07-168203>
76. Spardy N, Covella K, Cha E, *et al.*, 2009, Human papillomavirus 16 E7 oncoprotein attenuates DNA damage checkpoint control by increasing the proteolytic turnover of claspin. *Cancer Res*, 69(17): 7022–7029.
<https://doi.org/10.1158/0008-5472.CAN-09-0925>
77. Zaman MM, Nomura T, Takagi T, *et al.*, 2013, Ubiquitination-deubiquitination by the TRIM27-USP7 complex regulates tumor necrosis factor alpha-induced apoptosis. *Mol Cell Biol*, 33(24): 4971–4984.
<https://doi.org/10.1128/MCB.00465-13>
78. Yoshihara H, Fukushima T, Hakuno F, *et al.*, 2012, Insulin/insulin-like growth factor (IGF) stimulation abrogates an association between a deubiquitinating enzyme USP7 and insulin receptor substrates (IRSs) followed by proteasomal degradation of IRSs. *Biochem Biophys Res Commun*, 423(1): 122–127.
<https://doi.org/10.1016/j.bbrc.2012.05.093>
79. Sarkari F, Wheaton K, La Delfa A, *et al.*, 2013, Ubiquitin-specific protease 7 is a regulator of ubiquitin-conjugating enzyme Ube2E1. *J Biol Chem*, 288(23): 16975–16985.
<https://doi.org/10.1074/jbc.M113.469262>
80. Bhattacharya S, Ghosh MK, 2015, HAUSP regulates c-MYC expression via de-ubiquitination of TRRAP. *Cell Oncol (Dordr)*, 38(4): 265–277.
<https://doi.org/10.1007/s13402-015-0228-6>
81. Li N, Zhao Z, Liu P, *et al.*, 2021, Upregulation of deubiquitinase USP7 by transcription factor FOXO6 promotes EC progression via targeting the JMJD3/CLU axis. *Mol Ther Oncolytics*, 20: 583–595.
<https://doi.org/10.1016/j.omto.2020.12.008>
82. Episkopou H, Diman A, Claude E, *et al.*, 2019, TSPYL5 depletion induces specific death of ALT cells through USP7-dependent proteasomal degradation of POT1. *Mol Cell*, 75(3): 469–482.e466.
<https://doi.org/10.1016/j.molcel.2019.05.027>
83. Felle M, Joppien S, Németh A, *et al.*, 2011, The USP7/Dnmt1 complex stimulates the DNA methylation activity of Dnmt1 and regulates the stability of UHRF1. *Nucleic Acids Res*, 39(19): 8355–8365.
<https://doi.org/10.1093/nar/gkr528>
84. Zhu Q, Ding N, Wei S, *et al.*, 2020, USP7-mediated deubiquitination differentially regulates CSB but not UVSSA upon UV radiation-induced DNA damage. *Cell Cycle*, 19(1): 124–141.
<https://doi.org/10.1080/15384101.2019.1695996>
85. Lu X, Zhang Y, Zheng Y, *et al.*, 2021, The miRNA-15b/USP7/KDM6B axis engages in the initiation of osteoporosis by modulating osteoblast differentiation and autophagy. *J Cell Mol Med*, 25(4): 2069–2081.
<https://doi.org/10.1111/jcmm.16139>
86. Lee KW, Cho JG, Kim CM, *et al.*, 2013, Herpesvirus-associated ubiquitin-specific protease (HAUSP) modulates peroxisome proliferator-activated receptor γ (PPAR γ) stability through its deubiquitinating activity. *J Biol Chem*, 288(46): 32886–32896.
<https://doi.org/10.1074/jbc.M113.496331>
87. Salsman J, Jagannathan M, Paladino P, *et al.*, 2012, Proteomic profiling of the human cytomegalovirus UL35 gene products reveals a role for UL35 in the DNA repair response. *J Virol*, 86(2): 806–820.

- <https://doi.org/10.1128/JVI.05442-11>
88. Heimbucher T, Hunter T, 2015, The *C. elegans* ortholog of USP7 controls DAF-16 stability in insulin/IGF-1-like signaling. *Worm*, 4(4): e1103429.
<https://doi.org/10.1080/21624054.2015.1103429>
89. Jäger W, Santag S, Weidner-Glunde M, *et al.*, 2012, The ubiquitin-specific protease USP7 modulates the replication of Kaposi's sarcoma-associated herpesvirus latent episomal DNA. *J Virol*, 86(12): 6745–6757.
<https://doi.org/10.1128/JVI.06840-11>
90. Maertens GN, El Messaoudi-Aubert S, Elderkin S, *et al.*, 2010, Ubiquitin-specific proteases 7 and 11 modulate polycomb regulation of the INK4a tumour suppressor. *Embo J*, 29(15): 2553–2565.
<https://doi.org/10.1038/emboj.2010.129>
91. Sarasin A, 2012, UVSSA and USP7: New players regulating transcription-coupled nucleotide excision repair in human cells. *Genome Med*, 4(5): 44.
<https://doi.org/10.1186/gm343>
92. Frey Y, Franz-Wachtel M, Macek B, *et al.*, 2022, Proteasomal turnover of the RhoGAP tumor suppressor DLC1 is regulated by HECTD1 and USP7. *Sci Rep*, 12(1): 5036.
<https://doi.org/10.1038/s41598-022-05099-9>
93. Yi L, Cui Y, Xu Q, *et al.*, 2016, Stabilization of LSD1 by deubiquitinating enzyme USP7 promotes glioblastoma cell tumorigenesis and metastasis through suppression of the p53 signaling pathway. *Oncol Rep*, 36(5): 2935–2945.
<https://doi.org/10.3892/or.2016.5099>
94. Xiang Q, Ju H, Li Q, *et al.*, 2018, Human herpesvirus 8 interferon regulatory factors 1 and 3 mediate replication and latency activities via interactions with USP7 deubiquitinase. *J Virol*, 92(7): e02003–17.
<https://doi.org/10.1128/JVI.02003-17>
95. Du Z, Song J, Wang Y, *et al.*, 2010, DNMT1 stability is regulated by proteins coordinating deubiquitination and acetylation-driven ubiquitination. *Sci Signal*, 3(146): ra80.
<https://doi.org/10.1126/scisignal.2001462>
96. Bhattacharya S, Ghosh MK, 2014, HAUSP, a novel deubiquitinase for Rb MDM2 the critical regulator. *Febs J*, 281(13): 3061–3078.
<https://doi.org/10.1111/febs.12843>
97. Lee HR, Choi WC, Lee S, *et al.*, 2011, Bilateral inhibition of HAUSP deubiquitinase by a viral interferon regulatory factor protein. *Nat Struct Mol Biol*, 18(12): 1336–1344.
<https://doi.org/10.1038/nsmb.2142>
98. Ching W, Koyuncu E, Singh S, *et al.*, 2013, A ubiquitin-specific protease possesses a decisive role for adenovirus replication and oncogene-mediated transformation. *PLoS Pathog*, 9(3): e1003273.
<https://doi.org/10.1371/journal.ppat.1003273>
99. Yang P, Xie J, Li Y, *et al.*, 2020, Deubiquitinase USP7-mediated MCL-1 up-regulation enhances arsenic and benzo(a)pyrene co-exposure-induced cancer stem cell-like property and tumorigenesis. *Theranostics*, 10(20): 9050–9065.
<https://doi.org/10.7150/thno.47897>
100. Huang Q, Qin D, Pei D, *et al.*, 2022, UBE2O and USP7 co-regulate RECQL4 ubiquitinylation and homologous recombination-mediated DNA repair. *FASEB J*, 36(1): e22112.
<https://doi.org/10.1096/fj.202100974RRR>
101. Wu Y, Gu H, Bao Y, *et al.*, 2022, USP7 sustains an active epigenetic program via stabilizing MLL2 and WDR5 in diffuse large B-cell lymphoma. *Cell Biochem Funct*, 40(4): 379–390.
<https://doi.org/10.1002/cbf.3702>
102. Holowaty MN, Zeghouf M, Wu H, *et al.*, 2003, Protein profiling with Epstein-Barr nuclear antigen-1 reveals an interaction with the herpesvirus-associated ubiquitin-specific protease HAUSP/USP7. *J Biol Chem*, 278(32): 29987–29994.
<https://doi.org/10.1074/jbc.M303977200>
103. Meulmeester E, Pereg Y, Shiloh Y, *et al.*, 2005, ATM-mediated phosphorylations inhibit Mdmx/Mdm2 stabilization by HAUSP in favor of p53 activation. *Cell Cycle*, 4(9): 1166–1170.
<https://doi.org/10.4161/cc.4.9.1981>
104. de Bie P, Zaaroor-Regev D, Ciechanover A, 2010, Regulation of the Polycomb protein RING1B ubiquitination by USP7. *Biochem Biophys Res Commun*, 400(3): 389–395.
<https://doi.org/10.1016/j.bbrc.2010.08.082>
105. He J, Zhu Q, Wani G, *et al.*, 2014, Ubiquitin-specific protease 7 regulates nucleotide excision repair through deubiquitinating XPC protein and preventing XPC protein from undergoing ultraviolet light-induced and VCP/p97 protein-regulated proteolysis. *J Biol Chem*, 289(39): 27278–27289.
<https://doi.org/10.1074/jbc.M114.589812>
106. Xia X, Liao Y, Huang C, *et al.*, 2019, Deubiquitination and stabilization of estrogen receptor α by ubiquitin-specific protease 7 promotes breast tumorigenesis. *Cancer Lett*, 465: 118–128.
<https://doi.org/10.1016/j.canlet.2019.09.003>
107. Sun X, Ding Y, Zhan M, *et al.*, 2019, Usp7 regulates Hippo pathway through deubiquitinating the transcriptional coactivator Yorkie. *Nat Commun*, 10(1): 411.
<https://doi.org/10.1038/s41467-019-08334-7>
108. Lee JE, Park CM, Kim JH, 2020, USP7 deubiquitinates and stabilizes EZH2 in prostate cancer cells. *Genet Mol Biol*,

- 43(2): e20190338.
<https://doi.org/10.1590/1678-4685-GMB-2019-0338>
109. Khoronenkova SV, Dianov GL, 2013, USP7S-dependent inactivation of Mule regulates DNA damage signalling and repair. *Nucleic Acids Res*, 41(3): 1750–1756.
<https://doi.org/10.1093/nar/gks1359>
110. Zhu Q, Sharma N, He J, *et al.*, 2015, USP7 deubiquitinase promotes ubiquitin-dependent DNA damage signaling by stabilizing RNF168. *Cell Cycle*, 14(9): 1413–1425.
<https://doi.org/10.1080/15384101.2015.1007785>
111. Ni W, Lin S, Bian S, *et al.*, 2020, USP7 mediates pathological hepatic de novo lipogenesis through promoting stabilization and transcription of ZNF638. *Cell Death Dis*, 11(10): 843.
<https://doi.org/10.1172/JCI98765>
112. Franqui-Machin R, Hao M, Bai H, *et al.*, 2018, Destabilizing NEK2 overcomes resistance to proteasome inhibition in multiple myeloma. *J Clin Invest*, 128(7): 2877–2893.
<https://doi.org/10.1038/s41467-020-16038-6>
113. Ma P, Yang X, Kong Q, *et al.*, 2014, The ubiquitin ligase RNF220 enhances canonical Wnt signaling through USP7-mediated deubiquitination of β -catenin. *Mol Cell Biol*, 34(23): 4355–4366.
<https://doi.org/10.1128/MCB.00731-14>
114. Kim JM, Yang YS, Park KH, *et al.*, 2020, A RUNX2 stabilization pathway mediates physiologic and pathologic bone formation. *Nat Commun*, 11(1): 2289.
<https://doi.org/10.1038/s41467-020-16038-6>
115. Jang SY, Jang SW, Ko J, 2012, Regulation of ADP-ribosylation factor 4 expression by small leucine zipper protein and involvement in breast cancer cell migration. *Cancer Lett*, 314(2): 185–197.
<https://doi.org/10.1016/j.canlet.2011.09.028>
116. Hofseth LJ, Hussain SP, Harris CC, 2004, p53: 25 years after its discovery. *Trends Pharmacol Sci*, 25(4): 177–181.
<https://doi.org/10.1016/j.tips.2004.02.009>
117. Sabapathy K, Lane DP, 2018, Therapeutic targeting of p53: All mutants are equal, but some mutants are more equal than others. *Nat Rev Clin Oncol*, 15(1): 13–30.
<https://doi.org/10.1038/nrclinonc.2017.151>
118. Kruse JP, Gu W, 2009, Modes of p53 regulation. *Cell*, 137(4): 609–622.
<https://doi.org/10.1016/j.cell.2009.04.050>
119. Momand J, Zambetti GP, Olson DC, *et al.*, 1992, The mdm-2 oncogene product forms a complex with the p53 protein and inhibits p53-mediated transactivation. *Cell*, 69(7): 1237–1245.
[https://doi.org/10.1016/0092-8674\(92\)90644-r](https://doi.org/10.1016/0092-8674(92)90644-r)
120. Hu M, Gu L, Li M, *et al.*, 2006, Structural basis of competitive recognition of p53 and MDM2 by HAUSP/USP7: Implications for the regulation of the p53-MDM2 pathway. *PLoS Biol*, 4(2): e27.
<https://doi.org/10.1371/journal.pbio.0040027>
121. Sheng Y, Saridakis V, Sarkari F, *et al.*, 2006, Molecular recognition of p53 and MDM2 by USP7/HAUSP. *Nat Struct Mol Biol*, 13(3): 285–291.
<https://doi.org/10.1038/nsmb1067>
122. Ali A, Raja R, Farooqui SR, *et al.*, 2017, USP7 deubiquitinase controls HIV-1 production by stabilizing Tat protein. *Biochem J*, 474(10): 1653–1668.
<https://doi.org/10.1042/BCJ20160304>
123. Cummins JM, Rago C, Kohli M, *et al.*, 2004, Tumour suppression: Disruption of HAUSP gene stabilizes p53. *Nature*, 428(6982):1 p following 486.
<https://doi.org/10.1038/nature02501>
124. Gao L, Zhu D, Wang Q, *et al.*, 2021, Proteome analysis of USP7 substrates revealed its role in melanoma through PI3K/Akt/FOXO and AMPK pathways. *Front Oncol*, 11: 650165.
<https://doi.org/10.3389/fonc.2021.650165>
125. Yao Y, Zhang Y, Shi M, *et al.*, 2018, Blockade of deubiquitinase USP7 overcomes bortezomib resistance by suppressing NF- κ B signaling pathway in multiple myeloma. *J Leukoc Biol*, 104(6): 1105–1115.
<https://doi.org/10.1002/JLB.2A1017-420RR>
126. Cai J, Chen HY, Peng SJ, *et al.*, 2018, USP7-TRIM27 axis negatively modulates antiviral type I IFN signaling. *Faseb J*, 32(10): 5238–5249.
<https://doi.org/10.1096/fj.201700473RR>
127. Colleran A, Collins PE, O'Carroll C, *et al.*, 2013, Deubiquitination of NF- κ B by ubiquitin-specific protease-7 promotes transcription. *Proc Natl Acad Sci U S A*, 110(2): 618–623.
<https://doi.org/10.1073/pnas.1208446110>
128. Zeng M, Zhang X, Xing W, *et al.*, 2022, Cigarette smoke extract mediates cell premature senescence in chronic obstructive pulmonary disease patients by up-regulating USP7 to activate p300-p53/p21 pathway. *Toxicol Lett*, 359: 31–45.
<https://doi.org/10.1016/j.toxlet.2022.01.017>
129. Duan D, Shang M, Han Y, *et al.*, 2022, EZH2-CCF-cGAS Axis Promotes Breast Cancer Metastasis. *Int J Mol Sci*, 23(3): 1788.
<https://doi.org/10.3390/ijms23031788>
130. Qi SM, Cheng G, Cheng XD, *et al.*, 2020, Targeting USP7-mediated deubiquitination of MDM2/MDMX-p53 pathway for cancer therapy: Are we there yet? *Front Cell Dev Biol*, 8: 233.
<https://doi.org/10.3389/fcell.2020.00233>
131. Cheng X, Zhang B, Guo F, *et al.*, 2022, Deubiquitination

- of FBP1 by USP7 blocks FBP1-DNMT1 interaction and decreases the sensitivity of pancreatic cancer cells to PARP inhibitors. *Mol Oncol*, 16(7): 1591–1607.
<https://doi.org/10.1002/1878-0261.13149>
132. Zhang T, Periz G, Lu YN, *et al.*, 2020, USP7 regulates ALS-associated proteotoxicity and quality control through the NEDD4L-SMAD pathway. *Proc Natl Acad Sci U S A*, 117(45): 28114–28125.
<https://doi.org/10.1073/pnas.2014349117>
133. Hou R, Li Y, Luo X, *et al.*, 2022, ENKUR expression induced by chemically synthesized cinobufotalin suppresses malignant activities of hepatocellular carcinoma by modulating β -catenin/c-Jun/MYH9/USP7/c-Myc axis. *Int J Biol Sci*, 18(6): 2553–2567.
<https://doi.org/10.7150/ijbs.67476>
134. Tang LJ, Zhou YJ, Xiong XM, *et al.*, 2021, Ubiquitin-specific protease 7 promotes ferroptosis via activation of the p53/TfR1 pathway in the rat hearts after ischemia/reperfusion. *Free Radic Biol Med*, 162: 339–352.
<https://doi.org/10.1016/j.freeradbiomed.2020.10.307>
135. Yang X, Jin J, Yang J, *et al.*, 2021, Expression of ubiquitin-specific protease 7 in oral squamous cell carcinoma promotes tumor cell proliferation and invasion. *Genet Mol Biol*, 44(4): e20210058.
<https://doi.org/10.1590/1678-4685-GMB-2021-0058>
136. Li T, Guan J, Li S, *et al.*, 2014, HSCARG downregulates NF- κ B signaling by interacting with USP7 and inhibiting NEMO ubiquitination. *Cell Death Dis*, 5(5): e1229.
<https://doi.org/10.1038/cddis.2014.197>
137. Liu G, Liu Q, Yan B, *et al.*, 2020, USP7 inhibition alleviates H(2)O(2)-induced injury in chondrocytes via inhibiting NOX4/NLRP3 pathway. *Front Pharmacol*, 11: 617270.
<https://doi.org/10.3389/fphar.2020.617270>
138. Forand A, Koumakis E, Rousseau A, *et al.*, 2016, Disruption of the phosphate transporter Pit1 in hepatocytes improves glucose metabolism and insulin signaling by modulating the USP7/IRS1 interaction. *Cell Rep*, 16(10): 2736–2748.
<https://doi.org/10.1016/j.celrep.2016.08.012>
139. Li Q, Sun H, Luo D, *et al.*, 2021, Lnc-RP11-536K7.3/SOX2/HIF-1 α signaling axis regulates oxaliplatin resistance in patient-derived colorectal cancer organoids. *J Exp Clin Cancer Res*, 40(1): 348.
<https://doi.org/10.1186/s13046-021-02143-x>
140. Zhou Z, Yao X, Li S, *et al.*, 2015, Deubiquitination of Ci/Gli by Usp7/hausp regulates hedgehog signaling. *Dev Cell*, 34(1): 58–72.
<https://doi.org/10.1016/j.devcel.2015.05.016>
141. Wilson P, Abdelmotti L, Norcross R, *et al.*, 2021, The role of USP7 in the Shoc2-ERK1/2 signaling axis and Noonan-like syndrome with loose anagen hair. *J Cell Sci*, 134(21): jcs258922.
<https://doi.org/10.1242/jcs.258922>
142. von Locquenghien M, Rozalén C, Celià-Terrassa T, 2021, Interferons in cancer immunoeediting: Sculpting metastasis and immunotherapy response. *J Clin Invest*, 131(1): e143296.
<https://doi.org/10.1172/JCI143296>
143. Takaoka A, Yanai H, 2006, Interferon signalling network in innate defence. *Cell Microbiol*, 8(6): 907–922.
<https://doi.org/10.1111/j.1462-5822.2006.00716.x>
144. Chu TS, 1974, Interferon and its clinical application. *Zhonghua Yi Xue Za Zhi*, 9: 576–579.
145. Xu LG, Wang YY, Han KJ, *et al.*, 2005, VISA is an adapter protein required for virus-triggered IFN-beta signaling. *Mol Cell*, 19(6): 727–740.
<https://doi.org/10.1016/j.molcel.2005.08.014>
146. Fitzgerald KA, McWhirter SM, Faia KL, *et al.*, 2003, IKKepsilon and TBK1 are essential components of the IRF3 signaling pathway. *Nat Immunol*, 4(5): 491–496.
<https://doi.org/10.1038/ni921>
147. Yu CF, Peng WM, Schlee M, *et al.*, 2018, SOCS1 and SOCS3 target IRF7 degradation to suppress TLR7-mediated Type I IFN production of human plasmacytoid dendritic cells. *J Immunol*, 200(12): 4024–4035.
<https://doi.org/10.4049/jimmunol.1700510>
148. Blumer T, Coto-Llerena M, Duong FHT, *et al.*, 2017, SOCS1 is an inducible negative regulator of interferon λ (IFN- λ)-induced gene expression *in vivo*. *J Biol Chem*, 292(43): 17928–17938.
<https://doi.org/10.1074/jbc.M117.788877>
149. Ding L, Li J, Li W, *et al.*, 2018, p53 mediated IFN- β signaling to affect viral replication upon TGEV infection. *Vet Microbiol*, 227: 61–68.
<https://doi.org/10.1016/j.vetmic.2018.10.025>
150. Lee HR, Toth Z, Shin YC, *et al.*, 2009, Kaposi's sarcoma-associated herpesvirus viral interferon regulatory factor 4 targets MDM2 to deregulate the p53 tumor suppressor pathway. *J Virol*, 83(13): 6739–6747.
<https://doi.org/10.1128/JVI.02353-08>
151. Li X, Wang T, Tao Y, *et al.*, 2022, Inhibition of USP7 suppresses advanced glycation end-induced cell cycle arrest and senescence of human umbilical vein endothelial cells through ubiquitination of p53. *Acta Biochim Biophys Sin (Shanghai)*, 54(3): 311–320.
<https://doi.org/10.3724/abbs.2022003>
152. Mandras SA, Mehta HS, Vaidya A, 2020, Pulmonary hypertension: A brief guide for clinicians. *Mayo Clin Proc*,

- 95(9): 1978–1988.
<https://doi.org/10.1016/j.mayocp.2020.04.039>
153. Poch D, Mandel J, 2021, Pulmonary hypertension. *Ann Intern Med*, 174(4): ITC49–ITC64.
<https://doi.org/10.7326/AITC202104200>
154. Zhu Y, Zhang Q, Yan X, *et al.*, 2021, Ubiquitin-specific protease 7 mediates platelet-derived growth factor-induced pulmonary arterial smooth muscle cells proliferation. *Pulm Circ*, 11(4): 20458940211046131.
<https://doi.org/10.1177/20458940211046131>
155. Uhlen M, Zhang C, Lee S, *et al.*, 2017, A pathology atlas of the human cancer transcriptome. *Science*, 357(6352): eaan2507.
<https://doi.org/10.1126/science.aan2507>
156. Davoodpour P, Landström M, Welsh M, 2007, Reduced tumor growth *in vivo* and increased c-Abl activity in PC3 prostate cancer cells overexpressing the Shb adapter protein. *BMC Cancer*, 7: 161.
157. Qu CF, Chen TY, Wang YT, *et al.*, 2018, Primary prevention model of liver cancer in rural China. *Zhonghua Zhong Liu Za Zhi*, 40(7): 481–489.
<https://doi.org/10.3760/cma.j.issn.0253-3766.2018.07.001>
158. Dong L, Yu L, Bai C, *et al.*, 2018, USP27-mediated cyclin E stabilization drives cell cycle progression and hepatocellular tumorigenesis. *Oncogene*, 37(20): 2702–2713.
<https://doi.org/10.1038/s41388-018-0137-z>
159. Zhang W, Zhang J, Xu C, *et al.*, 2020, Ubiquitin-specific protease 7 is a drug-able target that promotes hepatocellular carcinoma and chemoresistance. *Cancer Cell Int*, 20: 28.
<https://doi.org/10.1186/s12935-020-1109-2>
160. Cai JB, Shi GM, Dong ZR, *et al.*, 2015, Ubiquitin-specific protease 7 accelerates p14(ARF) degradation by deubiquitinating thyroid hormone receptor-interacting protein 12 and promotes hepatocellular carcinoma progression. *Hepatology*, 61(5): 1603–1614.
<https://doi.org/10.1002/hep.27682>
161. Chauhan D, Tian Z, Nicholson B, *et al.*, 2012, A small molecule inhibitor of ubiquitin-specific protease-7 induces apoptosis in multiple myeloma cells and overcomes bortezomib resistance. *Cancer Cell*, 22(3): 345–358.
162. Colland F, Formstecher E, Jacq X, *et al.*, 2009, Small-molecule inhibitor of USP7/HAUSP ubiquitin protease stabilizes and activates p53 in cells. *Mol Cancer Ther*, 8(8): 2286–2295.
<https://doi.org/10.1158/1535-7163.MCT-09-0097>
163. Nicholson B, Suresh Kumar KG, 2011, The multifaceted roles of USP7: New therapeutic opportunities. *Cell Biochem Biophys*, 60(1–2): 61–68.
164. Reverdy C, Conrath S, Lopez R, *et al.*, 2012, Discovery of specific inhibitors of human USP7/HAUSP deubiquitinating enzyme. *Chem Biol*, 19(4): 467–477.
<https://doi.org/10.1016/j.chembiol.2012.02.007>
165. Altun M, Kramer HB, Willems LI, *et al.*, 2011, Activity-based chemical proteomics accelerates inhibitor development for deubiquitylating enzymes. *Chem Biol*, 18(11): 1401–1412.
<https://doi.org/10.1016/j.chembiol.2011.08.018>
166. Fan YH, Cheng J, Vasudevan SA, *et al.*, 2013, USP7 inhibitor P22077 inhibits neuroblastoma growth via inducing p53-mediated apoptosis. *Cell Death Dis*, 4(10): e867.
<https://doi.org/10.1038/cddis.2013.400>
167. Shin SB, Kim CH, Jang HR, 2020, Combination of inhibitors of USP7 and PLK1 has a strong synergism against paclitaxel resistance. *Int J Mol Sci*, 21(22): 8629.
<https://doi.org/10.3390/ijms21228629>
168. Fu C, Zhu X, Xu P, *et al.*, 2019, Pharmacological inhibition of USP7 promotes antitumor immunity and contributes to colon cancer therapy. *Oncol Targets Ther*, 12: 609–617.
<https://doi.org/10.2147/OTT.S182806>
169. Yang L, Cao N, Miao Y, *et al.*, 2021, Morin acts as a USP7 inhibitor to hold back the migration of rheumatoid arthritis fibroblast-like synoviocytes in a “Prickle1-mTORC2” dependent manner. *Mol Nutr Food Res*, 65(19): e2100367.
<https://doi.org/10.1002/mnfr.202100367>
170. Yu Z, Wei X, Liu L, *et al.*, 2022, Indirubin-3'-monoxime acts as proteasome inhibitor: Therapeutic application in multiple myeloma. *EBioMedicine*, 78: 103950.
<https://doi.org/10.1016/j.ebiom.2022.103950>
171. Hu T, Zhang J, Sha B, *et al.*, 2019, Targeting the overexpressed USP7 inhibits esophageal squamous cell carcinoma cell growth by inducing NOXA-mediated apoptosis. *Mol Carcinog*, 58(1): 42–54.
<https://doi.org/10.1002/mc.22905>
172. Becker K, Marchenko ND, Palacios G, *et al.*, 2008, A role of HAUSP in tumor suppression in a human colon carcinoma xenograft model. *Cell Cycle*, 7(9): 1205–1213.
<https://doi.org/10.4161/cc.7.9.5756>